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(54) Ubidecarenone-containing liposome and pharmaceutical composition containing the same.

(57) A ubidecarenone-containing liposome is provided which permits ubidecarenone to be transferred to a target organ at an increased speed. There is also disclosed a pharmaceutical composition containing the ubidecarenone-containing liposome acc. to the invention.

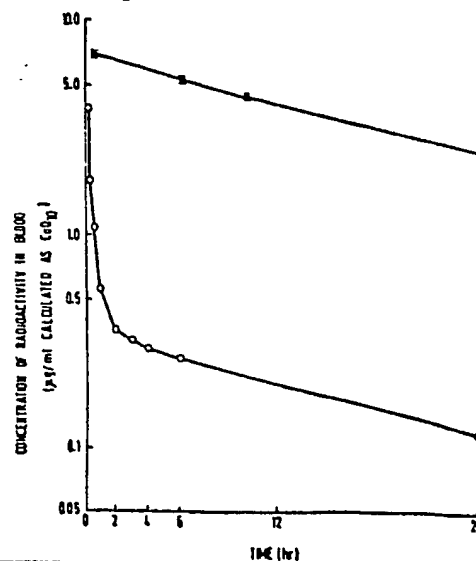


FIG. 1

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UBIDECARENONE-CONTAINING LIPOSOME
AND PHARMACEUTICAL COMPOSITION CONTAINING THE SAME

This invention relates to a ubidecarenone-containing liposome which permits rapid transfer of ubidecarenone from the blood to a target organ, as well as a pharmaceutical composition containing the same.

5 Ubidecarenone, also known as coenzyme Q₁₀, has recently come into widespread clinical use as a medicine effective for improving the function of the heart.

There is still room for improvement in the speed of transfer of this substance from the blood to a target organ
10 in intravenous or oral administration. Since the aforesaid substance is a medicine intended for improving the function of the heart, it is desired that the speed of its transfer to the target organ in the initial stage and the amount of its transfer be great. However, if this substance is formed into
15 a pharmaceutical preparation in accordance with a conventional technique, the speed of its disappearance from the blood is considerably low, and therefore the speed of its transfer to the target organ and its amount transferred are reduced.

Since ubidecarenone is a normally solid lipid-soluble
20 substance having a melting point of 48 to 52°C, it must be

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solubilized by a conventional technique involving the use of a surfactant such as HCO (polyoxyethylene-hardened castor oil) in order to render it convenient for intravenous administration.. However, if it is administered in the thus
5 solubilized state, the speed of disappearance of ubidecarenone from the blood is low, and the speed of its transfer in the initial stage to the heart, spleen and liver, i.e., the target organs, especially to the heart is low.

10

In view of the above background, the present inventors have made various investigations about a ubidecarenone-containing composition which is effective for increasing the transfer of ubidecarenone from the blood to predetermined target
15 organs. These investigations have led to the discovery that such a composition can be provided by incorporating ubidecarenone in liposome.

Thus, according to this invention, there is provided a ubidecarenone-containing liposome.

20

Figure 1 is a graphic representation showing the results obtained in Experimental Example 1 to be given hereinbelow,
25 and Figures 2 to 4 are graphic representations showing the results obtained in Experimental Example 2 to be given

hereinbelow.

5 Experimental Examples to be given hereinbelow show that
after administration of the liposome of this invention, the
speed of disappearance of ubidecarenone from the blood was
much higher, and the amount of ubidecarenone transferred
within a predetermined period to organs except the lungs
10 and kindeys was larger, as compared with the case of administering
the solubilized ubidecarenone prepared in accordance with the
conventional technique.

Generally, the liposome itself is known, and a conventional
method is available for its production. For the first time
15 in accordance with this invention, a ubidecarenone-containing
liposome has been produced by applying the liposome
technology to ubidecarenone, thereby solving the aforesaid
basic problem of ubidecarenone and greatly increasing the
utilitarian value of ubidecarenone.

20 In the liposome of this invention, ubidecarenone is
contained in a membrane constituting the liposome. The
liposome is composed basically of a phospholipid and a sterol.
Ubidecarenone is present together with these substances and
dispersed uniformly within the membrane.

25 Examples of the phospholipid used in this invention are
phosphatidyl choline, phosphatidyl ethanolamine, phosphatidyl

serine, sphingomyelin, dicetylphosphoric acid, stearylamine, phosphatidyl glycerol, phosphatidic acid, phosphatidyl inositol and mixtures thereof. It is however not specifically limited to these exemplified species. On the other hand, cholesterol is most preferred as the sterol.

The proportions of ubidecarenone, the phospholipid and the sterol in the liposome are preferably such that at least 10 moles of the phospholipid and at least 1 mole of the sterol are present per mole of ubidecarenone. For example, per mole of ubidecarenone, the proportion of the phospholipid is preferably 10 to 30 moles, if it is egg yolk phosphatidyl choline, and the proportion of the sterol is preferably 1 to 10 moles, if it is cholesterol.

Preferred proportions in moles of various phospholipids and cholesterol per mole of ubidecarenone are shown in the following table.

Examples		1	2	3	4	5	6	7
Phospholipid	PC	10	15	18	12	5	9	20
	DCP				2			5
	PS					5		
	SA						1	5
Cholesterol		5	5	4	4	5	5	5

PC : phosphatidyl choline

DCP: dicetylphosphoric acid

PS : phosphatidyl serine

SA : stearylamine

The liposome of this invention, when observed under an electron microscope, is a spherical particle having a particle diameter of about 0.1 to 5.0 μm . Its lyophilized product is aggregated and in appearance it is the form of a block. When
5 put into water or salt solutions, it disperses well to give a homogeneous solution.

The liposome of this invention can be produced substantially in accordance with the conventional technique for the production of liposome. For example, it can be produced by
10 charging a membrane-constituting component of the liposome and ubiquinone into an eggplant-shaped flask, adding chloroform to dissolve these substances, then evaporating the solvent, peeling the resulting membrane by adding a glass bead and a suitable buffer etc., ultrasonically treating the resulting solution, passing
15 the treated solution through a column of Sephadex or Sepharose, collecting liposome fractions, and removing the solvent. The removal of the solvent can be effected by, for example, lyophilization.

If the lyophilization is carried out for at least 3 hours
20 under a pressure of not more than 2.0 torr, the intended object of this invention can be achieved and the liposome can be taken out as a powder. The invention, however, is not limited to these specific conditions. As shown in a working example to be given hereinbelow, the lyophilization is desirably carried
25 out for 5 hours under a pressure of 0.3 torr, for example.

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The invention also comprises a pharmaceutical composition, which contains the ubidecarenone-containing liposome according to the invention besides usual pharmaceutically acceptable carriers and/or diluents.

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The following Experimental Examples illustrate the effect of the liposome of this invention.

Experimental Example 1

5 Sample:-

Liposome was prepared by the same method as described in Example 1 except that $^{14}\text{C-CoQ}_{10}$ was used instead of CoQ_{10} described in Example

1. The resulting liposome was used as an assay sample. A control sample was obtained by adding HCO-60 to $^{14}\text{C-CoQ}_{10}$, the amount of the former being four times that of the latter, and subjecting the solution to ultrasonication, and adding physiological saline to a concentration of 0.6 mg/ml.

Procedure:-

15 1. Animal experiment

The assay sample was injected in an amount of 0.6 mg as $^{14}\text{C-CoQ}_{10}$ /kg into the left femoral vein of male guinea pigs (body weight 300 - 350 g). The control sample was administered in the same dose by the same method. The animals were then left in a cage, and every predetermined period of time, blood was drawn from the ear vein. The concentration of $^{14}\text{C-CoQ}_{10}$ in the blood was measured by the following method.

2. Measurement of radioactivity in the blood

- 25 20 μl or 50 μl of blood was taken from the ear vein, and solubilized with 0.75 ml of Soluene 350/isopropyl alcohol (1/1). Several drops of aqueous hydrogen peroxide were added to decolorize the solution. Then, 5 ml of instagel/0.5N HCl (9/1) was added, and radioactivity was measured by means of a liquid scintillation counter.

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3. Results

Variations with time in the radioactivity concentration of $^{14}\text{C-CoQ}_{10}$ in the blood after the administration of the assay sample or the control sample are shown in Figure 1.

5 In Figure 1, the curve marked by \bigcirc refers to the case of administering the assay sample (average of two runs), and the curve marked by \square to the case of administering the control sample (an average of 3 runs, and a standard deviation). As shown in Figure 1, the disappearance of $^{14}\text{C-CoQ}_{10}$ in the assay sample from the blood is indicated by two straight lines which are bent at 1 hour after the administration. The half-life period of its disappearance is 11.5 minutes in the first phase and 15.6 hours in the second phase. The disappearance of $^{14}\text{C-CoQ}_{10}$ in the control sample from the blood is indicated by one straight line, and the half-life period of its disappearance is 20.1 hours. It is seen from these results that the disappearance of $^{14}\text{C-CoQ}_{10}$ in the assay sample from the blood is much more rapid than that of $^{14}\text{C-CoQ}_{10}$ in the control sample.

20 Experimental Example 2

Samples:-

The same assay sample and control sample as described in Experimental Example 1 were used.

25 Procedure:-

1. Animal experiment

Each of the above samples was injected in a dose of 0.6 mg/kg into the left femoral vein of male guinea pigs (body



weight 300 g - 350 g). The animals were then left in a cage. Thirty minutes and 24 hours respectively after the administration, the animals were killed by decapitation, and the organs were removed. In order to prevent contamination of the brain and heart by blood, physiological saline was circulated from the left ventricle to the jugular vein to draw the blood, and then the brain and heart were removed.

2. Measurement of radioactivity in the tissues

About 100 mg of each organ was added to 0.5 ml of Soluene 350, and incubated at 50°C for 2 hours to dissolve the tissues. Then, 6 ml of instagel/0.5N HCl (9/1) was added, and radioactivity was measured by using a liquid scintillation counter.

3. Results

Thirty minutes and 24 hours respectively after the intravenous injection of $^{14}\text{C-CoQ}_{10}$ in the assay sample and $^{14}\text{C-CoQ}_{10}$ in the control sample, the concentrations of radioactivity in the main organs were expressed in terms of CoQ_{10} , and the transfer of the former to the organs was compared with that of the latter. The results are shown in Figures 2 to 4.

In Figures 2 to 4, the hatched columns  refer to the case of administering the assay sample (an average of 2 runs), and the blank columns , to the case of administering the control sample (an average of 3 runs, and a standard deviation, provided that in the case of the results obtained 30 minutes

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after the administration with regard to the brain and heart, the indicated values are averages of 2 runs). In regard to the distribution of $^{14}\text{C-CoQ}_{10}$ to the tissues, its concentration increased more rapidly, and lasted longer, in the assay sample than in the control sample. Thus, the distribution of the assay sample differs from that of the control sample. In regard to the transfer of $^{14}\text{C-CoQ}_{10}$ to the brain, heart, liver, spleen and adrenal gland, it was distributed more rapidly in higher concentrations in the case of the assay sample than in the case of the control sample. It was specifically found that 30 minutes after the administration, $^{14}\text{C-CoQ}_{10}$ in the assay sample was distributed in the heart in a concentration 16 times as high as that in the control sample and in the brain 2 times as high as that in the control sample.

The following Examples illustrate the present invention further.

Example 1

36 mg (4.5×10^{-5} mole) of egg yolk phosphatidyl choline, 5.85 mg (1.5×10^{-5} mole) of cholesterol and 2.16 mg (2.5×10^{-6} mole) of ubiquinone, i.e. CoQ_{10} , were dissolved in 2 ml of chloroform. Chloroform was removed under reduced pressure by using a rotary evaporator. The resulting thin film was dried overnight by a desiccator under reduced pressure to remove chloroform completely. One glass bead and 3.0 ml of a buffer A (to be described below) were added twice to the thin

film left on the bottom of the desiccator, and the mixture was shaken by a vortex mixer until the film was completely peeled off.

5 The solution was then transferred to a branched test tube, and subjected to ultrasonication treatment at 42W for 53 minutes intermittently every 30 seconds by using a probe-type sonicator in an ice bath under a stream of argon to give a translucent pale yellow solution.

10 The solution was applied onto a column (1.6 x 45 cm) of Sepharose 4B, and eluted with the buffer A. Then, 1.8 ml portions of the eluate were taken respectively into 60 test tubes. A fraction which was eluted near the void volume

of this column was concentrated to a final volume of 1.7 ml (0.55 mg as CoQ_{10}) by means of a polycarbonate membrane having a pore size of 0.08 μm . The resulting liposome had a diameter of 80 to 100 μm by observation under an electron microscope.

The buffer A denotes a 0.01M phosphate buffer (pH 7.4) containing 0.1M NaCl.

20 Example 2

A 200 ml eggplant-shaped flask was charged with 60.0 mg (3 moles) of yolk phosphatidyl choline, 9.8 mg (1 mole) of cholesterol and 6.5 mg (0.3 moles) of ubidecarenone, and 4 ml of chloroform was added to dissolve them. The solvent was

evaporated under reduced pressure. Four milliliters of physiological saline and a glass bead were added, and dispersed by using a vortex mixer. The resulting multilayer film was transferred to a branched test tube, and under an atmosphere of nitrogen gas, subjected to ultrasonication treatment at 25 KW for 15 minutes in an ice bath. The solution was then charged onto a column of Sephadex G-50 and eluted with physiological saline as an eluent. Liposome fractions were collected and diluted to 30 ml. The diluted liposome fractions were lyophilized for 5 hours under a pressure of 0.3 torr.

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CLAIMS:

1. An ubidecarenone-containing liposome.
2. The liposome of claim 1 wherein ubidecarenone is present in the membrane of the liposome.
3. The liposome of claim 2 wherein the membrane of the
- 5 liposome is composed of a phospholipid and a sterol.
4. The liposome of claim 3 wherein the phospholipid is selected from the group consisting of phosphatidyl choline, phosphatidyl ethanolamine, phosphatidyl serine, sphingomyelin, dicetylphosphoric acid, stearylamine, phosphatidyl glycerol,
- 10 phosphatidic acid, phosphatidyl inositol and mixtures thereof.
5. The liposome of claim 3 or 4 wherein the sterol is cholesterol.
6. The liposome of any one of claims 3 to 5 which comprises 1 mole of ubidecarenone, at least 10 moles of the phospholipid, and at least 1 mole of the sterol.
- 15 7. Pharmaceutical composition, characterized in that it contains an ubidecarenone-containing liposome acc. for claims 1 to 6 as active substance besides usual pharmaceutically acceptable carriers and/or diluents.

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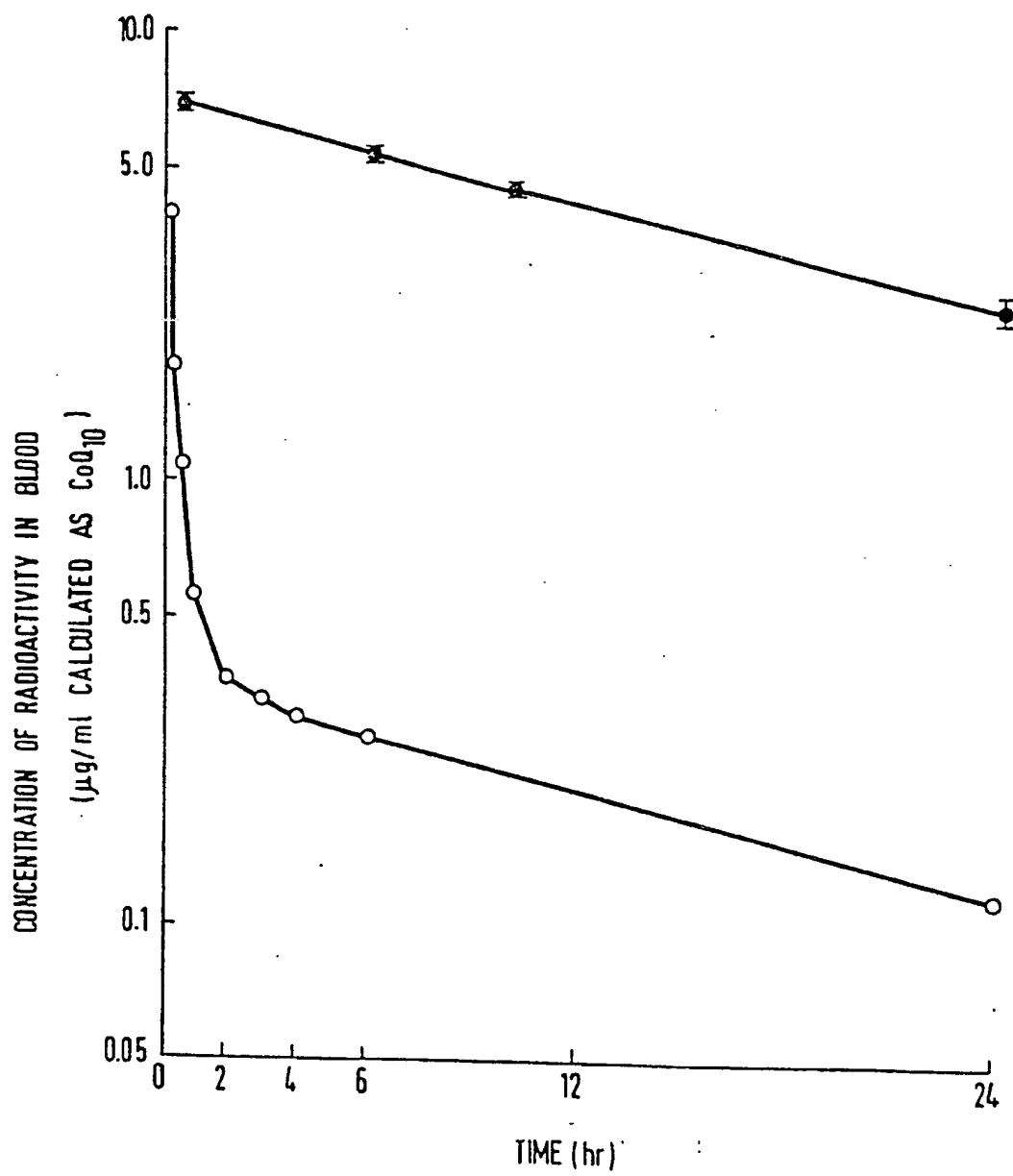


FIG . 1

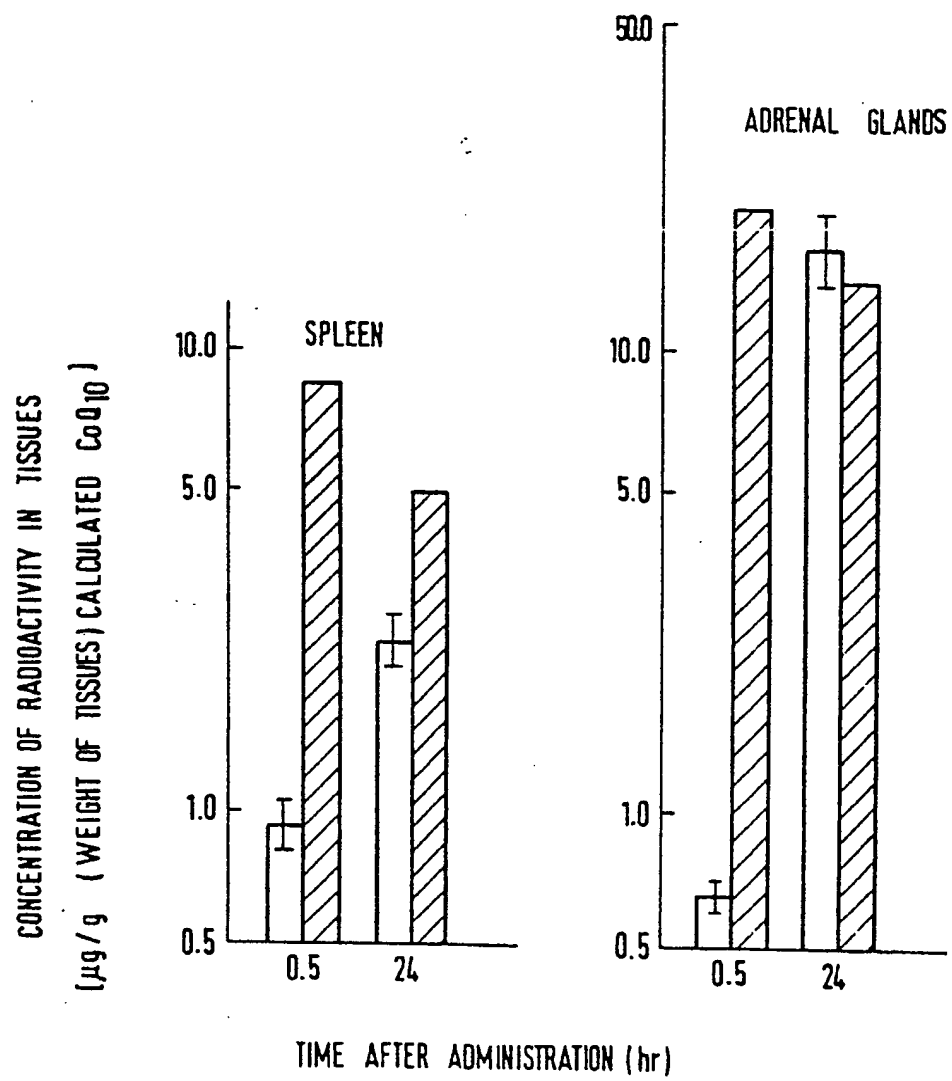


FIG. 2

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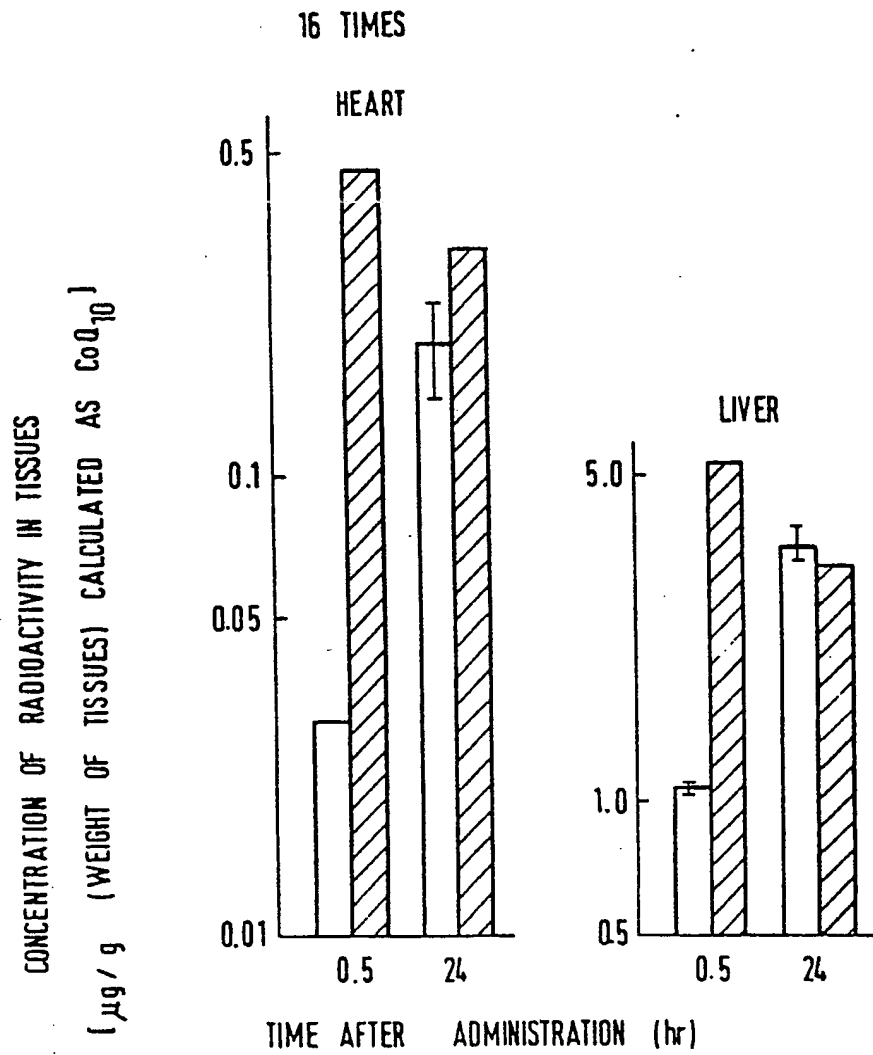


FIG. 3

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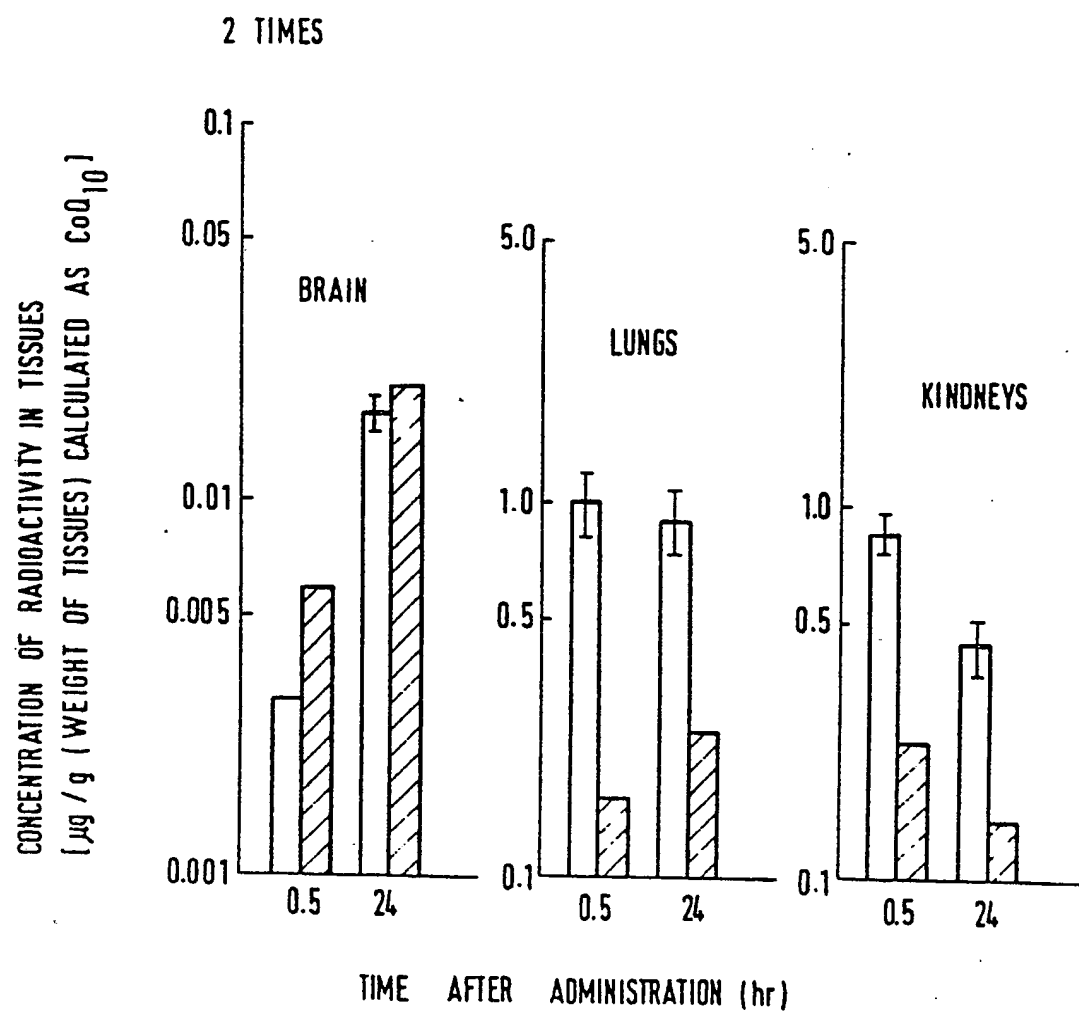


FIG. 4

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